Skeletal Structure of Neocopiamycin B from *Streptomyces hygroscopicus* var. crystallogenes

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Streptomyces hygroscopicus var. crystallogenes, IFM 1136 (FERM BP-576), produces antibiotic copiamycin (niphithricin, 2, Fig. 1)^{$1 \sim 4$}) along with minor macrocyclic lactone antibiotics, neocopiamycin A (3), demalonylcopiamycin (4), demalonylmethylcopiamycin (5) and guanidolide A, possessing a guanidino alkenyl side chain.^{5~7}) Copiamycin had been found to possess synergistic activity with chlorinated imidazole antimycotics such as clotrimazole, miconazole, econazole and especially ketokoazole.⁸⁾ ARAI et al. had reported that the strain produces a new macrocyclic lactone antibiotic, neocopiamycin B, when cobalt (II) chloride is added into the original medium.⁹⁾ Neocopiamycin B proved to be more active against some fungi than copiamycin (2) and/or neocopiamycin A (3) and low toxicity for mice (LD_{50}) : > 25 mg/kg, iv).⁹⁾ In this paper, we describe the skeletal structure of neocopiamycin B.

Materials and Methods

Instruments

FAB-MS spectrum was obtained with a Jeol DX303 mass spectrometer. MALDI-TOF MS spectrum was obtained with a PerSeptive Biosystems Voyager-DE STR with nitrogen laser (337 nm). Instrument operation was performed in delayed extraction reflector mode. The solid matrix used for MALDI was a-cyano-4-hydroxycinnamic acid. The milli-mass spectrum with MALDI-TOF MS was calibrated using angiotensin I and des-Arg(1)bradykinin as the internal standard: Number of data points was 50000 and the data were collected from 500 to 1550 mass range with 128 laser shots. Time interval between subsequent data points was 0.5 nsec. Accelerating voltage is 20 kV and grid voltage is 14.7 kV. NMR spectra were recorded on a Jeol JNM-α-500 (500 MHz for ¹H and 125 MHz for ¹³C) spectrometer at 50°C. As ¹H NMR chemical shift reference, the center signal of CD₃OD at δ 3.30 and DMSO- d_6 at δ 2.49, and as ¹³C NMR reference, the methyl signal of CD₃OD at δ 49.0 were used. ¹H NMR spectrum of 1 was assigned with COSY and HOHAHA (59 msec) spectra. ¹H and ¹³C NMR spectra of 2 were assigned with ¹H-¹H COSY, HOHAHA (35, 61, and 109 msec), J-resolved, HSQC, and ¹³C-¹H COSY spectra.

Purification of Neocopiamycin B (1) and Copiamycin (2)

Crude neocopiamycin B $(10 \text{ mg})^{9}$ was purified by preparative TLC (silica gel, 2-BuOH - H₂O - 28% ammonia (8:2:1)) to give pure 1 (2.5 mg). Purified copiamycin with preparative TLC (silica gel, 2-BuOH - H₂O



Fig. 1. Structures of copiamycin (2) and its derivatives $(3 \sim 5)$.

Copiamycin (2) $R_1 = COCH_2COOH, R_2 = H, R_3 = CH_3$ Neocopiamycin A (3) $R_1 = COCH_2COOH, R_2 = R_3 = H$ Demalonylcopiamycin (4) $R_1 = R_2 = H, R_3 = CH_3$ Demalonylmethylcopiamycin (5)

 $R_1 = H, R_2 = R_3 = CH_3$

	1	2	<u> </u>	2
С	δ _C	2	δ _H	
1	168.3	168.3		
2	123.2	123.2	5.87 (d J = 15.5)	5.86 (d. $J = 15.5$)
3	152.5	152.5	6.95 (dd, J = 8.5, 15.5)	6.93 (dd, J = 8.5, 15.5)
4	43.4	43.5	2.46 (m)	2.46 (m)
5	75.4	75.3	3.76	3.76 (td. $J = 5, 10$)
6	39.4	39.5	~ 1.50, ~ 1.75	~ 1.51, ~ 1.75
7	74.8	(75.0)*	3.72	~ 3.77
8	45.1	44.5	1.50 (m)	~ 1.55
9	72.6	(72.5)	3.75 (m)	~ 3.88
10	45.2	(44.7)		(~ 1.75)
11	37.0	(33.4)		(~ 1.58, ~ 1.65)
12	42.9	(44.5)		(~ 1.53)
13	33.4	(72.3)		~ 3.88
14	39.6	41.9		$\sim 1.67, \sim 1.78$
15	102.6	99.8		
16	76.1	77.3	3.75 - 3.9	3.35 (d, J = 9)
17	67,6	(69.8)	3.75 - 3.9	~ 3.87
18	42.8	41.2	~ 1.62, ~ 1.98	~ 1.29, ~ 1.90
19	65.8	65.9	4.00 (m)	4.09
20	42.1	42.1	~ 1.75, ~ 1.85	~ 1.67, ~ 1.78
21	71.4	71.1	5.29 (septet-like)	5.22 (septet-like)
22	40.9	40.6	~ 1.75, ~ 1.85	1.63 - 1.81
23	65.2	(65.8)	3.95 (m)	~ 3.88
24	43.0	43.1	~ 1.4, ~ 1.6	~ 1.39, ~ 1.60
25	69.3	(69.3)	4.18 (td, $J = 5$, 10)	4.13 (ddd, J = 2, 4.5, 10)
26	45.3	(45.4)	1.50 (m)	~ 1.50
27	76.1	(76.0)	3.88 (t, $J = 8$)	~ 3.88
28	134.5	134.7†	5.46	$5.47 (\mathrm{dd}, J = 8, 15.5)$
29	134.9	134.7†	5.55 (dd, J = 8, 15.5)	5.53 (dd, J = 8, 15.5)
30	39.6	39.7	2.54 (m)	2.52 (m)
31	80.9	80.4	4.73 (t, J = 6)	$4.75 (\mathrm{dd}, J = 5, 7)$
32	33,3	33.0	1.90 (m)	~ 1.92
33	42.3	(42.4)	~ 1.30	$(\sim 0.90, \sim 1.32)$
34	30.6	(30.8)	~ 1.60	(~ 1.58)
35	37.1	37.3		$\sim 1.09, \sim 1.28$
20	27.5	27.0	1.09 ()	$\sim 1.26, \sim 1.57$
3/	33.7	33,7	~ 1.90 (III) 5 48 (dd like $L = 6.15$)	~ 1.97 5 48
20	132,9	132.8	5.48 (dd - fixe, 5) = 0, 15) 5.43 (dt $I = 6, 15)$	5.43 5.42 (td $I = 6.15.5$)
39 40	30.3	30.6	2.08 (td $I = 6.7$)	2.07 (td I = 7.7)
40	20.5	20.0	$\sim 1.65 \text{ (m)}$	~ 1.65
42	42.0	42.0	3.17 (t. $J = 7$)	3.17 (t. $J = 7$)
43	16.5	16.5	1 12 (d J = 6.5)	1.12 (d. $J = 7$)
44	10.6	(10.6)	0.88 (d, J = 7)	0.89 (d, $J = 7$)
45	14.7	(14.7)	0.88 (d, J = 8)	(0.88, d, J = 7)
46	11.5	11.4	0.79 (d, J = 7)	0.79 (d, $J = 7$)
47	17.8	17.7	0.99(d, J = 7)	0.99 (d, $J = 7$)
48	15.3	(15.1)	0.87 (d, J = 7)	(0.88, d, J = 7)
49	20.8	(20.7)	0.92 (d, J = 7)	(0.87, d, J = 7)
50	158.4	158.4		
51	28.3	28.3	2.83 (s)	2.84 (s)
1'	171.5	171.6		
2'	(a)	(a)	(a)	(a)
3'	(b)	174.0		

Table 1. ${}^{13}C$ and ${}^{1}H$ NMR data of neocopiamycin B (1) and copiamycin (2) in CD₃OD.

* Signals show in parentheses are tentative assignments. [†] Appeared at δ 134.66 (correlated with the signal at δ 5.55) and 134.69 (correlated with signal at δ 5.46). (a): The signal(s) did not appear because of easily exchange of H₂-2' with deuterium at the measurement temperature (50°C), [1: δ 3.06 (2H, br s), **2**: δ 3.05 (2H, br s) in DMSO-d₆)]. (b): The signal was not observed because of a limitation in the quantity of the available sample.

(4:1)) was recrystallized from MeOH-H₂O.

Neocopiamycin B (1)

Neocopiamycin B (1) was obtained as a colorless powder. MALDI-TOF MS: m/z 1042.6802 (M+H)⁺, Δ +1.1 milli-mass unit (mmu) for C₅₄H₉₆N₃O₁₆, 1024.6681, $\Delta - 0.4 \,\mathrm{mmu}$ for $C_{54}H_{94}N_3O_{15}$, 956.6801, Δ + 1.3 mmu for C₅₁H₉₄N₃O₁₃, 938.6700, Δ + 1.9 mmu for $C_{51}H_{92}N_3O_{12}$, 920.6546, $\Delta - 2.9 \text{ mmu}$ for $C_{51}H_{90}$ - N_3O_{11} . FAB-MS: m/z 1042 $(M + H)^+$, 1024, 938, 448, 360, 348, 320, 294, 282, 266, 252, 224, 210, 182, 168, 154, 100, 87, 73, 57. ¹H NMR (DMSO- d_6): δ 0.68 (3H, d, J = 7 Hz, H₃-46), 0.72 (3H, d, J = 7 Hz, H₃-49), 0.76 (3H, d, J = 6.5 Hz, H₃-48), 0.80 (3H, d, J = 7 Hz, H₃-45), 0.85 $(3H, d, J=7 Hz, H_3-44), \sim 0.86 (H-33), 0.87 (3H, d,$ $J = 6.5 \text{ Hz}, \text{ H}_3\text{-}43$), 1.06 (3H, d, $J = 7 \text{ Hz}, \text{ H}_3\text{-}47$), ~1.22, ~1.30 (H-33), ~1.32 (H-34), ~1.42 (H-26), ~1.51 (H-12), ~ 1.52 (H₂-41), ~ 1.53 (H-8), ~ 1.62 , ~ 1.81 $(H_2-20 \text{ and/or } H_2-22), \sim 1.77 (H-32), \sim 1.93 (H_2-37),$ 1.98 (2H, H₂-40), 2.46 (1H, br, H-4), 2.54 (1H, br, H-30), 2.70 (3H, s, H_3 -51), 2.95 (1H, d, J = 14 Hz, H-16), 3.05 (3H, br, H-17 and H₂-42), \sim 3.50, \sim 1.27, ~3.80 (HOCHCH₍₂₎CHOH), ~3.52, ~1.33, ~3.72 (HOCHCH₍₂₎CHOH), ~3.58 (H-19), ~3.79 (H-27), 3.98 (1H, br d, J = 10 Hz, H-25), 4.60 (1H, dd, J = 5and 8 Hz, H-31), 5.04 (1H, m, H-20), 5.33~5.43 (3H, m, H-28, -38, and -39), 5.48 (1H, dd, J=7 and 15 Hz, H-29), 5.89 (1H, d, J = 16 Hz, H-2), 6.85 (1H, dd, J = 9and 16 Hz, H-3).

Results and Discussion

Crude neocopiamycin B^{9} was purified by preparative TLC to give pure neocopiamycin B (1). Previously the molecular formula of 1 was reported as C₅₃H₉₁O₁₇N₃. 4H₂O by its FAB-MS and elemental analysis.⁹⁾ But, matrix-assisted laser desorption/ionization time-of-flight mass spectrum (MALDI-TOF MS) of 1 gave a MW, m/z1042.6802 $(M+H)^+$, which fits the molecular formula, $C_{54}H_{95}O_{16}N_3.$ The mass spectrum also showed the fragment ions at m/z 1024.6681 (MH-H₂O)⁺, 956.6801 $[MH - (malonyl - H)]^+$, 938.6700 $(M - O - malonyl)^+$, and 920.6546 (938 - H₂O) indicating that the compound is a malonyl ester. The other significant ion was not observed in the MALDI-TOF MS. The ¹H and ¹³C NMR spectra of 1 (Table 1) showed the presence of the alkenyl side chain and macrocyclic moieties consist with eight methyl groups (one is substituted with a nitrogen atom; $\delta_{\rm C}$ 28.3), six olefinic carbons, one sp^2 carbon

Fig. 2. Partial structures $(A \sim D)$ of neocopiamycin B (1).



bearing to nitrogen atoms ($\delta_{\rm C}$ 158.4), one carbonyl carbon ($\delta_{\rm C}$ 168.3), one hemiketal carbon ($\delta_{\rm C}$ 102.6) and 34 methine or methylene carbons in which 11 carbons are substituted with an oxygen function ($\delta_{\rm C}$ 65.2~80.9). The spin-spin network founded with the ¹H-¹H COSY spectrum (in CD₃OD) showed four partial structures, $A \sim D$ (Fig. 2). By the comparison of chemical shifts and coupling patterns of 1 with those of copiamycin (2), three partial structures A, C, and D could be assigned as $C1 \sim C9$, $C18 \sim C34$, and $C37 \sim C42$, respectively. The 21-H signal of copiamycin and its derivatives shows a significant coupling pattern (septet-like signal). The 21-H signal of 1 appeared at δ 5.29 (septet-like signal) indicating that the malonyl group links to the C21 position (the signal of desmalonyl compounds, 4 and 5, appears at δ 4.04 and 4.05, respectively, and that of malonylated compounds, 2 and 3, appears at δ 5.22 and 5.21, respectively).^{$5 \sim 7$} These partial structures were supported further by the ¹³C NMR spectra: On the comparison of ¹³C NMR spectra of 1 and 2, the chemical shifts of the assignable signals for the partial structures of 1 resembled to those of 2 (Table 1). On the other hand, the FAB-MS of 1 gave the fragment ions at m/z 100, 154, 168, 182, 210, 224 and 254 derived from cleavages on the guanidino alkenyl side chain (Fig. 3) as well as that of copiamycin.⁵⁾ Thus, the compound 1 has the same alkenyl side chain as 2, 4 and 5. These above data suggested that neocopiamycin B may be 13-deoxy or 16-deoxy or 17-deoxy copiamycin. The 13-deoxy or 17-deoxy structure for 1 was suggested by the chemical shifts of C15 of 1 (δ 102.6)



Fig. 3. Fragmentations in the positive FAB-MS of neocopiamycin B (1).

as following. The signals of $2 \sim 4$ appeared more upfield (about 3 ppm, δ 99.3 ~ 99.8) than that of **1**. The upfield shift could be explain by the γ -trans effect of a hydroxy group (C13-OH or C17-OH).¹⁰⁾ Furthermore, if compound 1 is 16-deoxycopiamycin, the C15 signal must be observed at near δ 90 with a release from β effect of C16-OH (downfield shift of $8 \sim 10 \text{ ppm}$).¹⁰⁾ Thus, the 16-deoxy structure was eliminated. The discrimination between the remained two structures was established by the ¹H NMR spectrum of 1 measured in DMSO- d_6 . In the solvent, the spectrum was observed as broad signals, but hydroxy methine proton signals appeared in relatively wide range ($\delta 2.95 \sim 4.00$). The H-16 signal was appeared at δ 2.95 as a doublet (J = 14 Hz) correlated to the signals at δ 3.05 (H-17 and H₂-42) in the ¹H-¹H COSY spectrum indicating the presence of the C16-OH and C17-OH groups. From the above all data, the skeletal structure of neocopiamycin B was assigned to 13-deoxycopiamycin (1, Fig. 4).

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References

- ARAI, T.; S. KURODA, H. OHARA, Y. KATO & H. KAJI: Copiamycin, a new antifungal antibiotic-derived from *S. hygroscopicus* var. *crystallogenes*. J. Antibiotics, Ser. A 18: 63~67, 1965
- FUKUSHIMA, K.; T. ARAI, S. IWASAKI, M. NAMIKOSHI & S. OKUDA: Studies on macrocyclic lactone antibiotics. VI. Skeletal structure of copiamycin. J. Antibiotics 35: 1480~1494, 1982
- BASSI, L.; B. JOOS, P. GASSMANN, H.-P. KAISER, H. LEUENBERGER & W. KELLER-SCHIERLEIN: Stoffwechselprodukte von Mikroorganismen, 218. Mitteilung. Versuche zur Strukturaufklärung von Niphimycin, 1. Teil. Reinigung und Charakterisierung der Niphimycine Iα und Iβ sowei Abbau mit Salpetersäure. Helv. Chim. Acta 66: 92~117, 1983
- GASSMANN, P.; L. HAGMANN, W. KELLER-SCHIERLEIN & D. SAMAIN: Stoffwechselprodukte von Mikroorganismen, 226. Mitteilung. Versuche zur Strukturanfklärung von Niphimycin, 3. Teil. Identität von Scopafungin mit Niphimycin I und Lage des Malonylrestes in Niphimycin und Copiamycin. Helv. Chim. Acta 67: 696~705, 1984
- ARAI, T.; J. UNO, I. HORIMI & K. FUKUSHIMA: Isolation of neocopiamycin A from *Streptomyces hygroscopicus* var. *crystallogenes*, the copiamycin source. J. Antibiotics 37: 103~109, 1984
- 6) FUKAI, T.; T. NOMURA, J. UNO & T. ARAI: Demalonylcopiamycin, a new antibiotic produced by *Streptomyces hygroscopicus* var. *crystallogenes*, the copiamycin source. Heterocycles 24: 3351~3358, 1986
- 7) FUKAI, T.; C. TAKAHASHI, T. NOMURA, J. UNO & T. ARAI: Guanidolide A, a novel antibiotic produced by *Streptomyces hygroscopicus* var. *crystallogenes*, the copiamycin source. Heterocycles 27: 2333~2340, 1988

- UNO, J.; M. L. SHIGEMATSU & T. ARAI: Novel synergism of two antifungal agents, copiamycin and imidazole. Antimicrob. Agents Chemother. 24: 552~ 559, 1983
- 9) ARAI, T.; I. HORIMI, A. IWAYAMA, K, NAKAMURA & Y. IWAMOTO: Neocopiamycin B manufacture with

Streptomyces. Jpn. Kokai Tokkyo Koho JP 62-158298, 1987; Chem. Abstr. 108: 110834, 1988

 WEHRLI, F. W.; A. P. MARCHAND & S. WEHRLI: Interpretation of Carbon-13 NMR Spectra, 2nd ed. pp. 51~54, John Wiley & Sons, Chichester, 1988